

Evaluation of Oxidative Stress during Toxoplasmosis in Pregnant Women

Sahib J. Abdul-Rahman¹, Omaima I. Mahmood² and Reem S. Abdul-Aziz³

¹Department of Biology, Faculty of Science;² Department of Microbiology, Faculty of Veterinary Medicine;³ Department of Biology, Faculty of Science
University of Tikrit, Iraq

Abstract: *Toxoplasma gondii* (*T. gondii*) is the causative agent of toxoplasmosis. It infects up to one third of the human population. This study has conducted in Tikrit Teaching Hospital from September 2013 until March 2014, to investigated the effect of infection with toxoplasmosis, in pregnant women, on some enzymatic and non enzymatic antioxidants. Seventy six blood samples were collected from infected women their ages between (16-46) years, divided into two groups (16-26) and (27-46) years, and also 25 blood samples from non-infected pregnant women as a control group and Superoxide dismutase (SOD), catalase (CAT) and Glutathione Peroxidase (G-Px), Glutathione (GSH), Ceruloplasmin (Cp), Uric acid and Albumin were determined. The results showed a significant increase in concentration of SOD and uric acid compared with control group while significant decrease in the concentration of CAT, G-Px, and GSH, while non significant differences in the concentration of albumin compared with the control group.

Keywords: *Toxoplasmosis, pregnant women, enzymatic and non enzymatic antioxidants*

1. Introduction

Toxoplasmosis is one of the common parasitic infections in tropical and subtropical climates. Its causative agent is *Toxoplasma gondii* (*T. gondii*). It exists in a chronic asymptomatic form in 500 million to 1 billion people. [1,2] *T. gondii* is widespread protozoan Parasite, that infects most types of warm-blooded mammals and causes opportunistic disease in humans. [3] *T. gondii* is an obligate intracellular parasite replicating inside a parasitophorous vacuole in a broad range of host cells including macrophages. Human infection may be acquired in several ways: (i) ingestion of undercooked infected meat containing *Toxoplasma* cysts; (ii) ingestion of the oocyst from focally contaminated hands or food; (iii) blood transfusion; (iv) transplacental transmission. [4,5] Toxoplasmosis can cause serious pathologies including hepatitis, pneumonia, blindness and severe neurological disorders. [6]

Toxoplasmosis may cause severe disorders in immunocompromised patients and in Pregnant women, because of high risk of transplacental transmission and the occurrence of multiple congenital lesions in the fetus. [7]

During infection, the immune effector cells are able to kill or inhibit its intracellular growth. This antiprotozoan activity produces a number of toxic products such reactive oxygen intermediates. While, within the host cell, *T. gondii* itself produces oxidants as by products of normal metabolism. Reactive oxygen species (ROS) are potentially destructive, capable of oxidizing proteins or lipids and causing chemical modifications to nucleic acids. [8] It is well documented that, under normal physiological conditions, an estimated 1–3% of inspired oxygen is converted to superoxide radicals and H₂O₂. The existence and development of cells in an oxygen containing environment would not be possible without the presence of a complicated defense system that includes enzymatic and nonenzymatic antioxidant components. [9]

The main objective of this study was to estimate some antioxidants level like enzymatic antioxidants: Superoxide dismutase (SOD), Catalase (CAT) and Glutathione Peroxidase (G-Px), and non-enzymatic antioxidants: Glutathione (GSH), Ceruloplasmin (Cp), Uric acid and Albumin, in serum of positive toxoplasmosis pregnant women. Results were compared to those obtained with uninfected pregnant women (control), for a better understanding of toxoplasmosis pathogenesis.

2. Materials and methods

Seventy six blood samples were collected from seropositive- Toxoplasmosis women and from (25) seronegative Toxoplasmosis women (Control group). Age range: 16-46 years old and who had been referred to Tikrit hospital in Salaheldin from September 2013 until March 2014. None of the participants of this study took medication or supplementation upon entering the study.

Serum was separated immediately and stored at -20°C until biochemical analysis. Enzyme linked immunosorbent assay (ELISA) test was performed on all of the samples using immunoglobulin G (IgG) Kit (Human, German) and the final results were recorded by ELISA reader (optical absorbance, OD = 450).

SOD activity was determined in erythrocyte hemolysates according to the method described by Sun et al. [10] This method provides that the rate of nitroblue tetrazolium (NBT) reduction to blue formazan by the superoxide anion generated by the xanthine oxidase (XOD) reaction is monitored spectrophotometrically at 560 nm. One unit of SOD was considered a 50% inhibition of reduction of NBT under the condition of the assay. The results were expressed as U/g Hb.

Glutathione peroxidase activity in serum was assayed colorimetrically, a method used by Nelson and Kulkarni. [11]

Serum glutathione was measured by a modified procedure utilizing Ellman's reagent and determined from a standard curve and expressed as nmol/mg protein. [11]

Catalase activity was measured spectrophotometrically at 240 nm by calculating the rate of degradation of H_2O_2 as the substrate of the enzyme using the Aebi method. [12] A molar absorption of $43.6 \text{ M}^{-1}\text{cm}^{-1}$ was used to determine catalase activity. Enzymatic activity was expressed as U/mg protein, one unit (U) of which was equal to 1 mole of H_2O_2 degraded/min/mg of protein.

Ceruloplasmin analysis was conducted by a spectrophotometric method, which included P-phenyldiamine dichloride (PPD) use.[13]

Uric acid concentration in plasma is determined by use of uricase to allantoin and hydrogen peroxide ($2\text{H}_2\text{O}_2$). [14]

Albumin Serum was determined by dye-binding method [15] using kit manufactured by bioMerieux. The measurement of albumin is based on its quantitative binding at pH 4.2 with bromocresol green (BCG) to form a blue-green complex.

Statistical Analysis:

The results are expressed as mean \pm SD. Our data were analyzed statistically using one-way analysis of variance. Group differences were determined using Duncan multiple range test. Statistical significance was considered at $p < 0.05$. [16]

3. Results

The results in table (1) revealed a significant increase ($P < 0.05$) in serum SOD activity between toxoplasmosis women and control group, while CAT activity and GSH-px decreased significantly ($P < 0.05$) in serum of Toxoplasmosis woman, compared with control group as shown in table (1) .

In the present study GSG and CP activity decreased significantly in sero positive Toxoplasmosis woman in compared to the control group. On the other hand, infection of the pregnant women with toxoplasmosis produced a significant increase in uric acid concentration in comparison with the healthy pregnant women. There was no significant difference in serum albumin between control and infected women as shown in table (2).

TABLE I: Enzymatic Antioxidants Level in Toxoplasmosis pregnant women

Categories	parameters		
	SOD U/gHb	Catalase k / gHb	GSH-px umol/L
Infected women (16-26 years)	669.4 \pm 249 a	0.1 \pm 0.13 c	0.8 \pm 0.1 c
Control group	248 \pm 19 b	0.1 \pm 0.31 a	1.78 \pm 0.4 b
Infected women (27-46 years)	852 \pm 160 a	0.11 \pm 0.1812 c	0.84 \pm 0.118 c
Control group	235.75 \pm 13 b	0.21 \pm 0.075 b	0.84 \pm 0.118 a

*Different letterers refer to significant differences at ($P < 0.05$)

TABLE II: Non enzymatic Antioxidants Level in Toxoplasmosis pregnant women

Categories	Parameters			
	CP (mg/dl)	Glutathion (Umol/L)	Albumin (g/dL)	Uric acid(mg/ml)
Infected women(16-26 years)	41.1 ± 298.5 d	1 ± 5.1 b	1.09 ± 5.3 a	0.7 ± 7.3 b
Control group	72.9 ± 481 a	0.9 ± 9.2 a	0.408 ± 4 a	0.1 ± 6.6 c
Infected women(27-46 years)	20.9 ± 354.5 c	0.9 ± 5.3 b	5.2 ± 6.2 a	1.2 ± 7.7 a
Control group	44.7 ± 441.6 b	1.2 ± 9.3 a	0.4 ± 4.2 a	0.4 ± 6.8 c

*Different letterers refer to significant differences at (P<0.05)

4. Discussion

Superoxide dismutase is an important physiological antioxidant defense mechanism in aerobic organism. This enzyme prevents the formation of the hydroxyl radical by detoxifying hydrogen peroxide. [17] In this study, SOD activity in pregnant women of the case group was higher, suggesting that elevation of these antioxidant enzyme provides mainly protection against ROS-induced tissue injury also neutrophils and macrophages release ROS as part of the oxidative burst during *T. gondii* infection.[18] ROS generation is controlled by the cellular antioxidant enzymes such as SOD which detoxifies superoxide to hydrogen peroxide (H₂O₂).¹⁸ this result agree with[19,20], although others pointed there was no changes in serum SOD activity in infected women with *T. gondii*., that related to increasing the severity of parasitemia and oxidative stress.[21,22]

ROS generation is controlled by the cellular antioxidant enzymes such as catalase which converts H₂O₂ to H₂O.[23] this result agree with[20,24]

The increased SOD activity was associated with a significant decrease in catalase activity in women of the case group leading to the accumulation of H₂O₂, which may be the cause of the induction of oxidative stress. [23]

Glutathione is the most abundant non-protein thiol source in the cell, which acts as a substrate for several enzymes, including glutathione peroxidase and GST and serves multiple functions in protecting tissues from oxidative damage and keeping the intracellular environment in the reduced state.[17,23] A significant depletion of glutathione and glutathione peroxidase activity were noted in the present study in serum of women infected with *T. gondii* which was the result of high oxidative stress and both antioxidants over-use by the cells. Moreover, the low glutathione levels, especially in the infected pregnant women with acute phase of toxoplasmosis, represent a decreased detoxicating capacity of pregnant [25] The decreased in level in serum of toxoplasmosis patients has been demonstrated [20,21,26]

The present study showed no significant differences in albumin ratio in seropositive Toxoplasmosis woman and this result is consistent with the of previous study.[23] But Boothroyd *et al.*[27] observed that toxoplasmosis led to an increase in serum protein and globulin concentrations and a decrease in serum albumin concentrations during the acute stage and decrease of albumin in the acute stage which indicates decrease in protein metabolism or increase catabolism. [28]

Ceruloplasmin is an acute phase protein that responses mildly to inflammation and tissue damage .[29] Some researchers pointed out that there are positive correlations between acute phase reactants and the severity of lesions as well as prognosis.[30] In the present study, CP concentration was lower in *T. gondii* seropositive pregnant women (P<0.05).It acts as an antioxidant through either prevention of decompartmentized iron acting as free radical catalysis or by directly inactivating the free radical and CP was far more effective as a peroxy radical scavenger than SOD, deferoxamine and BSA, but slightly less effective than catalase.[31] This result agreed with other researches.[32]

In positive toxoplasmosis group, the result of uric acid was significantly increased.[33] Uric acid plays different roles in human body and plays a role as an endogenous antioxidant. Uric acid is able to react with different free radicals forming relatively stable urate radical and thus stopping radical reactions. So it is a powerful antioxidant and is a scavenger of singlet oxygen and radicals.[8,33]

5. Acknowledgment

This study was supported by Department of Biology, Faculty of Science. University of Tikrit, Iraq.

6. References

- [1] j. Abdi, S. Shojaee, and A. Mirzaee , "Seroprevalence of toxoplasmosis in pregnant women in Islam province, Iran," *Iran J Parasitol*,3:34-7,. 2008.
- [2] N. Jalalou, M. Bandepour, and H. Khazan, "Recombinant SAG1 Antigen to detect *Toxoplasma gondii* specific immunoglobulin G in human sera by ELISA Test, " *Iran J Parasitol*, 5:1-9, 2010.
- [3] J. Dubey, S. Chunlei, K.Daniel, K. Howe, and S. David, "Identification of quantitative trait loci controlling acute virulence in *Toxoplasma gondii*", *Am Vet Med Assoc*,205,1593-1598,1994.
- [4] I.J. Blader, and J.P. Saeij, "Communication between *Toxoplasma gondii* and its host: impact on parasite growth, development, immune evasion, and virulence, " *APMIS*, vol.117,pp. 458-476, 2009.
<http://dx.doi.org/10.1111/j.1600-0463.2009.02453.x>
- [5] J. Montoya, and O. Liesenfeld , "Toxoplasmosis", *Lancet* , vol.363, pp.1965- 1976, 2004.
[http://dx.doi.org/10.1016/S0140-6736\(04\)16412-X](http://dx.doi.org/10.1016/S0140-6736(04)16412-X)
- [6] U. Karaman, T. Celik, and T.R. Kiran, "Malondialdehyde , glutathione ,and nitric oxide levels in *Toxoplasma gondii* Seropositive Patients," *Korean J Parasitol*,vol.46,pp.293-5, 2008.
<http://dx.doi.org/10.3347/kjp.2008.46.4.293>
- [7] W. Foulon, I. Villena, and B.Stray-petersen, "Treatment of toxoplasmosis during pregnancy a multicenter study of impact on fetal transmission and children's sequelae at age 1 year," *Am J Obstet Gyneco*, vol. 180, pp.410-415,1999.
[http://dx.doi.org/10.1016/S0002-9378\(99\)70224-3](http://dx.doi.org/10.1016/S0002-9378(99)70224-3)
- [8] B. Halliwell, and J.M. Gutteridge, *Antioxidant defenses in Free radicals in biology and medicine*, 4th ed. Oxford University Press, New York,1999, pp. 105 – 245.
- [9] C. Guohua, and L.Ronald, " Postrandial increase in serum antioxidant capacity in older women," *J Appl Physiol* , vol.89, pp. 877 – 883,2000.
- [10] Y. Sun, L.W. Oberley, Y. Li, "A simple method for clinical assay of superoxide dismutase," *Clin Chem*, vol.34, pp.497-500,1988.
- [11] J.L. Nelson, and A.P. Kulkarni, "Partial purification and characterization of a peroxidase activity from human placenta," *Biochem J*, vol.268: pp.739-747,1990.
<http://dx.doi.org/10.1042/bj2680739>
- [12] H. Aebi, " Catalase in vitro," *Methods Enzymol*,vol.105, pp.121-6, 1984.
[http://dx.doi.org/10.1016/S0076-6879\(84\)05016-3](http://dx.doi.org/10.1016/S0076-6879(84)05016-3)
- [13] J.P. Colombo,and R. Richterich, "Zur Bestimmung Des Caeruloplasmin im plasma," *Schweiz Med Wochen*, vol. 94, pp.715-720, 1964.
- [14] N.W. Tietz, *Textbook of clinical chemistry*, 3rd ed. C.A. Burtis; E.R. Ashwood, W.B. Saunders, 1999,pp: 819-861,1245-1250.
- [15] B.T. Doumas, W.A. Waston, and H.G. Bigg, "Albumin standards and the measurement of serum albumin with BCG," *Clin Chim Acta*, vol.31, pp. 87-96,1971.
[http://dx.doi.org/10.1016/0009-8981\(71\)90365-2](http://dx.doi.org/10.1016/0009-8981(71)90365-2)
- [16] R.G.D. Steel, and J.H. Torrie, *Principles and Procedures of Statistics*, 2nd ed. 1980, New York: McGraw-Hill Book Company, Inc. 1960, pp.87-80, 107-109, 125-127.
- [17] B. Halliwell, "Free radicals and antioxidants: Updating a personal view," *Nutr Rev*, vol. 70, pp. 257-65, 2012.
<http://dx.doi.org/10.1111/j.1753-4887.2012.00476.x>
- [18] G. Correa, C. Marques da Silva, A.C. de Abreu Moreira-Souza, "Activation of the P2X (7) receptor triggers the elimination of *Toxoplasma gondii* tachyzoites from infected macrophages," *Microbes Infect*,vol.12, pp.497-504, 2010.
<http://dx.doi.org/10.1016/j.micinf.2010.03.004>
- [19] A. j. Kaasch, and K. A. Joiner, "Targeting andsubcellular localization of *Toxoplasma gondii* catalase. Identification of peroxisomes in an apicomplexan parasite," *J Biol Chem*, vol.275,pp.1112 – 1118,2000.
<http://dx.doi.org/10.1074/jbc.275.2.1112>
- [20] J. Mahvash , S. Maryam, S. Shahnaz , A. Laila, and T. Fatemeh, "Evaluation of gender-related differences in response to oxidative stress in *Toxoplasma gondii* positive serum, " *Annal Milit & Health Scien Research* , vol. 12, pp. 64-69, 2014.

- [21] E.M. Al-Khshab, "Some antioxidants level in seropositive toxoplasmosis woman in Mosul, " *Tikrit J Pre Sci*, vol.15, pp.17-22, 2010.
- [22] A. Nurgül , Ç. Miyase, G. Bayram, K. Ruhi , N.G. Aycan, T. Hasan, and C. Sıla, "Evaluation of oxidative stress, hematological and biochemical parameters during *Toxoplasma gondii* infection in gerbils, " *Ankara Üniv Vet Fak Derg*, vol.62, pp.165-170, 2015.
http://dx.doi.org/10.1501/Vetfak_0000002675
- [23] M. Jafari, M. Salehi, H. Zardooz, " Response of liver antioxidant defense system to acute and chronic physical and psychological stresses in male rats, " *EXCLI J*, vol.13, pp.161-71, 2014.
- [24] M. Gilca, I. Stoian, D. Lixandru, L. Gaman, B. Virgolici, and V. Atanasiu, " Protection of erythrocyte membrane against oxidative damage by geriforte in healthy human subjects," *Rom J Intern Med*, vol. 47(3), pp.289-95, 2009.
- [25] X. Xu, T. Liu, A. Zhang, "Reactive oxygen species triggered trophoblast apoptosis is initiated by endoplasmic reticulum stress via activation of caspase-12, CHOP, and the JNK pathway in *Toxoplasma gondii* infection in Mice," *Infect Immun*, vol.80, pp.2121-32, 2012.
<http://dx.doi.org/10.1128/IAI.06295-11>
- [26] N. A. Bassam A. H. Farah, U. K. Wasan, "Effect of *Toxoplasma gondi* Infestation on Lipid Peroxidation and Certain Antioxidants in Pregnant Women in Mosul City," *Raf Jour Sci*, Vol.17, No.10 Biology, Special Issue, pp.16-25, 2006.
- [27] J. Boothroyd, M. Black, S. Bonnefoy, A. Hehl, I. Manger, and S. Tomavo, "Genetic and biochemical analysis of development in *Toxoplasma gondii*," *Phil Trans R Scotland*, vol.352, pp.1347-1354 , 1997.
<http://dx.doi.org/10.1098/rstb.1997.0119>
- [28] B. Esmaelnejad, M. Tavassoli, and S. Asri-Rezaei, "Investigation of hematological and biochemical parameters in small ruminants naturally infected with *Babesia ovis*, " *Vet Res Forum*, vol.3, pp. 31-36, 2012.
- [29] O. Gungor, B. Sunar, F. Ozcelik, Z. Aktas, S.S. Gokmen, "Serum sialic acid levels in acute myocardial infarction and relationship to ceruloplasmin," *Turk J Biochem*, vol. 29, pp. 226- 231, 2004.
- [30] C.L. Chen, F.T. Tang, H.C. Chen, C.Y. Chung, and M.K. Wong, "Brain lesion size and location: effects on motor recovery and functional outcome in stroke patients," *Arch Phys Med Rehabil*, vol. 81, pp. 447-452, 2000.
<http://dx.doi.org/10.1053/mr.2000.3837>
- [31] J.M.C. Gutteridge, " Ceruloplasmin: a plasma protein, enzymes, and antioxidant," *Ann Clin Biolchem*, vol.15, pp.293, 1978.
<http://dx.doi.org/10.1177/000456327801500170>
- [32] M.A. Nada, Y.Z. Yaha, K.A. Hussein, and T.H. Huda, "Serum Copper, Zinc, and Magnesium in *Toxoplasma* Seropositive women with a History of Abortion," *National J Chem*, , Vol. 20, pp. 595- 602, 2005.
- [33] M. Machin, M.F. Simoyi, K.P. Blemings, and H. Klandrof, (March 2004). Increased dietary protein elevates plasma uric acid associated with decreased oxidative stress in rapidly-growing broilers. *Comp Biochem Physiol B Biochem Mol Biol.[online]* 137. pp.282-290. available <http://www.ncbi.nlm.nih.gov/pubmed/15050525>.
<http://dx.doi.org/10.1016/j.cbpc.2004.01.002>